

Notes on Maxwell & Delaney

PSY710

11 single-factor within-subject designs

The ideas of blocking and the analysis of covariance were discussed in chapter 9. By explicitly incorporating individual differences among subjects into the design and analysis of experiments, blocking and ancova can significantly reduce error variance and, consequently, increase the power of our statistical tests. One can think of **within-subjects designs** as the ultimate extension of the blocking approach: each block consists of a single subject. If certain assumptions are met, this type of design can significantly increase the sensitivity of an experiment.

Up to now we have discussed experimental designs in which different treatments have been administered to different groups of subjects. Such designs are called *between-subjects designs*. In a within-subject design, each subject receives *all* treatments. This type of design is sometimes referred to as a *repeated-measures design*, because multiple measures are obtained on each subject. However, some statisticians prefer to use repeated-measures to refer only to designs in which the same dependent measure is taken at multiple times.

There are, of course, a wide range of within-subjects experimental designs. I will focus on designs in which the multiple measures obtained from each subject are all of the same type. I will *not* address designs that obtain measures that differ qualitatively. For example, we could imagine a situation where each subject provides dependent measures of response time and response accuracy in a visual detection task. Such data should be analyzed using a multivariate approach described in chapters 13 and 14 in your textbook, and will not be considered here.

The basic approach we will take to analyze within-subject data is to treat subjects as an experimental factor. However, there are several aspects of within-subjects designs that make the analysis more difficult. The first is that, in most within-subject designs, there is only one observation per cell (i.e., subject-treatment combination). That is to say, each subject is tested once, and only once, in each experimental condition. This means that we cannot measure “within-cell” error as we did in factorial experiments. The second distinguishing characteristic of this design is that the subjects factor is a **random factor**. Most experimental treatments are **fixed factors**: if the experiment was repeated, we would use the same levels on the experimental variables. Subjects, however, is a random factor: If the experiment was repeated, we would use different “levels” on the subjects factor (i.e., we

would get different subjects). A random factor adds variability to our measurements and, as we shall see, alters the analysis. Finally, unlike in previous designs, the observations in within-subjects designs are correlated, not independent. In other words, the residuals of our model are likely to have structure, and such structure will affect our analysis.

One more thing. Traditional methods for analyzing within-subject designs — the methods described here — are applicable only to *balanced* designs. In some ways, this is a major limitation of this approach. For example, the requirement of balanced data means that we must discard all of the data from a subject even if we lack a measurement in only one condition. Nevertheless, all of the following analyses assume that the design is balanced. Also, we will only consider designs with a single, fixed within-subject factor.

11.1 linear models

We start with the model

$$Y_{ij} = \mu + \alpha_j + \pi_i + (\pi\alpha)_{ij} + \epsilon_{ij} \quad (1)$$

where Y_{ij} is the score from subject i in condition j , μ is the intercept, α_j is the effect associated with condition j , π_i is the effect associated with subject i , $(\pi\alpha)_{ij}$ is the effect of the interaction between subject i and condition j , and ϵ_{ij} is the error for subject i in condition j .

We have a problem. The model in Equation 1 has too many parameters. Consider an experiment that has $n = 8$ subjects and $a = 4$ treatments for a total of 32 observations. Equation 1 includes an intercept (i.e., μ), $3 = a - 1$ treatment effects, $7 = n - 1$ subject effects, and $21 = (a-1)(n-1)$ interaction effects, which add up to a total of $1+3+7+21 = 32$ free parameters, which equals the number of observations. Therefore, the error terms in Equation 1 will be zero. The problem is that we have included an interaction term in our model, but there is no way to estimate that parameter from our data. Stated another way, there is no way to determine if the difference between the observation in each cell and the prediction based on the treatment and subject effects is due to an interaction or error. Therefore, I will simplify the model by dropping the interaction term

$$Y_{ij} = \mu + \alpha_j + \pi_i + \epsilon_{ij} \quad (2)$$

This interaction-less model is the full model for a design that consists of a single within-subjects factor and that has only a single measurement per cell. Note that the interaction effects do not disappear. Instead, they are incorporated into the error term (i.e., the residuals). By doing this, we are in some sense equating the *treatment* \times *subjects* interaction and error. I will return to this point below.

The null hypothesis being tested is

$$H_0 : \alpha_1 = \alpha_2 = \dots = \alpha_j = 0 \quad (3)$$

so a restricted model is

$$Y_{ij} = \mu + \pi_i + \epsilon_{ij} \quad (4)$$

The F test is computed the usual way

$$F = \frac{(E_R - E_F)/(df_R - df_F)}{E_F/df_F} \quad (5)$$

where E_F and E_R are $SS_{residuals}$ from the full and reduced models, respectively. The degrees of freedom for the residuals in the full model are worth examining. The degrees of freedom equal the number of observations minus the number of parameters in the model. You can verify that $df_F = (n - 1)(a - 1)$. Thus, $df_{residuals}$ for the model in Equation 2 equals the degrees of freedom for the (deleted) *treatment* \times *subjects* interaction term. Once again, we see that there is a connection between the *treatment* \times *subjects* interaction and error. The degrees of freedom for the reduced model is $n(a - 1)$, so $df_R - df_F = a - 1$.

11.2 model coefficients

When the model in Equation 2 is fit to data, the best-fitting (least-squares) coefficients are:

$$\begin{aligned} \hat{\mu} &= \bar{Y}_{..} \\ \hat{\alpha}_j &= \bar{Y}_{.j} - \bar{Y}_{..} \\ \hat{\pi}_i &= \bar{Y}_{i.} - \bar{Y}_{..} \end{aligned}$$

The intercept, μ , is the mean of all scores. The effect of treatment j (α_j) is the mean of the scores in treatment j , averaged across all subjects, minus the grand mean. The effect of subject i (π_i) is the mean score of subject i , averaged across treatments, minus the grand mean.

11.3 expected mean squares

Consider, again, Equation 2. Let's imagine that we fit this model to many sets of data and calculated the *average* parameter values. It can be shown¹ that the average, or expected, values of the parameters are the ones listed in Table 1. Notice that the expected value of $MS_{residuals}$ is the sum of error variance and a term related to the *treatment* \times *subjects* interaction. This result should not be surprising, because Equation 2 was created essentially by folding the interaction term into error. It is important for you to realize that the residuals and the interaction term are perfectly confounded in this design: The residual term *is* the interaction and *vice versa*. For this reason, some statistics packages label the residual term as *Treatment* \times *Subjects*.

¹See Kirk (1995) for a derivation of the expected mean squares.

Table 1: Expected Mean Squares for a design that has one, fixed within-subjects factor (**treatment**).

Effect	Type	$E(\text{Mean Square})$
treatment	fixed	$\sigma_e^2 + \sigma_{\pi\alpha}^2 + n \sum_{j=1}^a \alpha_j^2 / (a - 1)$
subjects	random	$\sigma_e^2 + a\sigma_{\pi}^2$
residuals		$\sigma_e^2 + \sigma_{\pi\alpha}^2$

Table 1 shows that $MS_{\text{treatment}}$ also is influenced by the interaction term. Because the interaction contributes equally to $MS_{\text{treatment}}$ and $MS_{\text{residuals}}$, it is reasonable to conclude that $\sigma_{\alpha}^2 > 0$ when $MS_{\text{treatment}} > MS_{\text{residuals}}$. In other words, a comparison of the models shown in Equations 2 and 4 provides a reasonable test of the null hypothesis. Notice, however, it is *not* reasonable to evaluate the effect of subject by dividing MS_{subjects} by $MS_{\text{residuals}}$. In fact, there is no unambiguous test of the effect of subjects. This lack of a test is not really a problem, however, because rarely are we interested in showing that subjects differ beyond what is expected by chance.

11.4 sphericity

A one-way, between-subjects ANOVA assumes that error variance is constant across conditions. A similar assumption is made in the one-way within-subjects ANOVA: Specifically, the assumption is that the error variances for all of the dependent variables are equal. Another fundamental assumption in the between-subjects ANOVA is that the errors – i.e., the ϵ_{ijk} 's – are independent. This assumption is reasonable in between-subjects designs that randomly assign subjects to conditions, but it is less reasonable in within-subjects studies. Indeed, it is reasonable to expect that errors for a given subject will be correlated, to some degree, across conditions. Therefore, the independent-errors assumption needs to be relaxed if we are to conduct a reasonable analysis of data collected in within-subjects experiments. Instead of assuming independence, we will assume that the errors exhibit a specific form of dependency, or correlation. In particular, the assumption is that all of the covariances² between dependent variables are equal. This combination of assumptions – equal variances for all dependent variables, and equal covariances between each pair of dependent variables – is known as the assumption of **compound symmetry**. The F calculated in Equation 5 is distributed as an F statistic with $df = [(a - 1), (n - 1)(a - 1)]$ if the dependent variables exhibit compound symmetry³.

²Given random variables X and Y with expected values $E(X)$ and $E(Y)$, the covariance of X and Y is $E(XY) - E(X)E(Y)$.

³Note that the same assumption is made for data collected in between-subjects designs, except that the standard assumption is that the covariances are all zero.

Although compound symmetry is sufficient for the F to be distributed correctly, it is not a necessary condition. Huynh and Feldt (1970) and Rouanet and Lépine (1970) showed that the F value is distributed as an F statistic if the variances of all *differences* among the dependent variables have the same variance:

$$\sigma_{Y_j - Y_k}^2 = \sigma_j^2 + \sigma_k^2 - 2\sigma_{jk} \quad (j \neq k) \quad (6)$$

where σ_{jk} is the covariance between dependent measures j and k . When this condition is met, the variance-covariance matrix of the dependent variables is said to be spherical, so this assumption is known as the **sphericity assumption**. It is important to remember that the sphericity assumption applies to all tests of a within-subject factor. Also, **the sphericity assumption is necessarily true whenever the F test for the within-subject factor has one degree of freedom in the numerator**.

Unfortunately, the sphericity assumption often is not valid. In such cases, the F value calculated in Equation 5 will not be distributed as F with the expected degrees of freedom. However, it will be distributed *approximately* as an F statistic with lower degrees of freedom (Box, 1954). Therefore, one strategy for dealing with violations of the sphericity assumption is to adjust our degrees of freedom before evaluating F . The modified degrees of freedom are $\epsilon(a - 1)$ and $\epsilon(n - 1)(a - 1)$, where ϵ is a number indicating the degree to which the sphericity assumption is violated. When sphericity exists, $\epsilon = 1$; otherwise, ϵ is less than one with a minimum value of $1/(a - 1)$.

In practice, of course, ϵ is not known and so must be estimated from the data. One strategy is to simply assume that ϵ is at its minimum value. When $\epsilon = 1/(a - 1)$, $df_1 = 1$ and $df_2 = (n - 1)$, and a within-subjects F test using these degrees of freedom is referred to as the Geisser-Greenhouse **conservative F test**, or as using the **lower-bound adjustment** of the degrees of freedom (Geisser and Greenhouse, 1958). Alternatively, we can derive numerical estimates of ϵ from the variance-covariance matrix of our dependent measures. Geisser and Greenhouse proposed one such estimate, $\hat{\epsilon}$. The degrees of freedom for the **adjusted F test** are $df_1 = \hat{\epsilon}(a - 1)$ and $df_2 = \hat{\epsilon}(n - 1)(a - 1)$. The Geisser-Greenhouse adjustment controls Type I error but is more powerful than the lower-bound adjustment. It is, however, slightly conservative as the true population ϵ approaches one, and therefore Huynh and Feldt (1976) proposed a different adjustment based on their estimate of epsilon denoted as $\tilde{\epsilon}$. The Huynh-Feldt procedure yields adjusted degrees of freedom $df_1 = \tilde{\epsilon}(a - 1)$ and $df_2 = \tilde{\epsilon}(n - 1)(a - 1)$. It, too, is more powerful than the lower-bound adjustment, and it is slightly less conservative than the Geisser-Greenhouse adjustment. Both the Geisser-Greenhouse and Huynh-Feldt adjustments are acceptable procedures, although the former probably does a slightly better job at controlling Type I error rates. The conservative F test, as its name implies, is the most conservative test available: if the conservative F test is significant, than tests based on $\hat{\epsilon}$ and $\tilde{\epsilon}$ will be significant, too.

11.5 R example

In this section I will analyze the data presented in Table 11.5 in your textbook. The data are from a fictitious experiment that measured cognitive ability in 12 children at 30, 36, 42, and 48 months of age. First, I read the data file and get it into the correct format.

```
> mw115<-read.table("chapter_11_table_5.dat")
> mw115$subj<-factor(x=1:12,labels="s",ordered=FALSE)
> names(mw115) <- c("age.30","age.36","age.42","age.48","subj")
> mw115
```

	age.30	age.36	age.42	age.48	subj
1	108	96	110	122	s1
2	103	117	127	133	s2
3	96	107	106	107	s3
4	84	85	92	99	s4
5	118	125	125	116	s5
6	110	107	96	91	s6
7	129	128	123	128	s7
8	90	84	101	113	s8
9	84	104	100	88	s9
10	96	100	103	105	s10
11	105	114	105	112	s11
12	113	117	132	130	s12

The data frame contains four measures taken on each of 12 subjects. To analyze these data, we first use `lm()` to create a **multivariate model** of the between-subjects effects. The four dependent variables – `age.30`, `age.36`, `age.42` and `age.48` – are combined into a single matrix using the `cbind()` command. There is no between-subjects variable in this experiment, so the model contains only an intercept, which in R is represented as 1:

```
> # multivariate linear model:
> mw115.mlm <- lm(cbind(age.30,age.36,age.42,age.48) ~ 1, data=mw115) # only intercept in this case
```

Now I have to inform R about the nature of the within-subjects design. First, I create a factor that contains the levels within-subjects variable:

```
> (age<-factor(x= c("a30","a36","a42","a48"),ordered=FALSE) )
```

```
[1] a30 a36 a42 a48
Levels: a30 a36 a42 a48
```

Note that the order of the levels is the same as the order of my model's multivariate dependent variable. Next, I have to specify the within-subjects design with a formula of the form $Y \sim myFactor$, or $Y \sim A + B + A : B$. There is only one within-subjects variable, *age*, so the appropriate formula for the current experiment is $Y \sim age$. Finally, I use the

Anova command in the `car` package to convert our multivariate R object, `mw115.mlm`, into an anova object. In the next command, note how I pass information about the within-subjects factor in the `idata` parameter, and the within-subjects design in the `idesign` parameter:

```
> #install.packages("car")
> library(car)
> mw115.aov <- Anova(mw115.mlm, idata=data.frame(age), idesign=~age, type="III")
```

Information about the within-subjects design was specified by a *one-sided* model formula, `~ age`. The `type` parameter is used to tell `Anova` to calculate Type III sums of squares. The fact that our data are balanced (and that we do not have a between-subjects factor) means that Type II and III sums of squares will be equivalent, and therefore the `type` parameter is superfluous in this case. Finally, we can construct an anova table with the `summary` command. Setting `multivariate` to false suppresses the print out of multivariate tests:

```
> summary(mw115.aov, multivariate=FALSE)
```

Univariate Type III Repeated-Measures ANOVA Assuming Sphericity

	SS	num Df	Error SS	den Df	F	Pr(>F)
(Intercept)	559872	1	6624	11	929.7391	5.586e-12 ***
age	552	3	2006	33	3.0269	0.04322 *

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Mauchly Tests for Sphericity

	Test statistic	p-value
age	0.24265	0.017718

Greenhouse-Geisser and Huynh-Feldt Corrections
for Departure from Sphericity

	GG eps	Pr(>F[GG])
age	0.60954	0.07479 .

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

	HF eps	Pr(>F[HF])
age	0.7248502	0.06353773

Several pieces of information are printed in the summary. The first part shows the standard ANOVA table: the p value for `age` is the one calculated if we assume that the

sphericity assumption is valid. The second part shows the results of the Mauchly test for sphericity: the significant p value indicates that the deviation from sphericity is significant. The Mauchly test has been criticized for having low power, and therefore you might want to use a liberal Type I error rate (e.g., $\alpha = 0.1$) when using it to evaluate sphericity. The final part of the output show $\hat{\epsilon}$ and $\tilde{\epsilon}$, as well as the corrected p values for the test of a main effect of `age`. Note that neither p value is significant. In a paper, you could report this result as follows:

[At the beginning of your Results section...] Statistical analyses were done with R (R Development Core Team, 2008). For within-subject tests of more than 1 degree-of-freedom, the Huynh-Feldt estimate of sphericity ($\tilde{\epsilon}$) was used to adjust p values of F tests conducted on within-subject variables (Maxwell and Delaney, 2004). [And later, when reporting the result of this analysis...] The effect of Age was not significant, $F(3, 33) = 3.027$, $\tilde{\epsilon} = 0.72$, $p = 0.063$.

11.5.1 using aov and Error

The ANOVA table produced by `Anova` does not include information about subjects. Normally this is not a problem because there is no F test that can be done to evaluate the effect of subject. However, the complete table is useful on those occasions when we want to compute the variance component for subjects. The following code shows how to produce such a table using the `aov` command. To perform the analysis, we first must convert the data from a wide-format matrix, in which each row contains all of the data from a single subject, to a long-format data frame, in which each row contains a *single* dependent variable:

```
> # store dependent vars in wide-format matrix:
> dat.mat <- with(mw115, cbind(age.30, age.36, age.42, age.48) )
> dat.mat[1:4,]

      age.30 age.36 age.42 age.48
[1,]    108    96    110    122
[2,]    103    117    127    133
[3,]     96    107    106    107
[4,]     84     85     92     99

> dim(dat.mat) # 48 data points in 12 rows

[1] 12  4

> # convert wide-form matrix to long-form data frame:
> dat.stack <- stack(data.frame(dat.mat))
> names(dat.stack) <- c("y", "age") # name columns
> subject <- factor(x=rep(1:12, times=4), labels="s") # create subject factor
```

```
> # combine everything:
> dat.long.form <- data.frame(dat.stack,subject)
> dim(dat.long.form)
```

```
[1] 48 3
```

```
> # 48 data points in 48 rows
> # each row contains ONE dependent var:
```

```
      y    age subject
1  108 age.30     s1
2  103 age.30     s2
3   96 age.30     s3
4   84 age.30     s4
```

```
[1] "..."
```

```
      y    age subject
13  96 age.36     s1
14 117 age.36     s2
15 107 age.36     s3
16  85 age.36     s4
```

```
[1] "..."
```

```
      y    age subject
25 110 age.42     s1
26 127 age.42     s2
27 106 age.42     s3
28  92 age.42     s4
```

```
[1] "..."
```

```
      y    age subject
37 122 age.48     s1
38 133 age.48     s2
39 107 age.48     s3
40  99 age.48     s4
```

Note how the data from a single subject is spread over multiple rows, and that the new `subject` factor indicates which subject contributed the data on each row. Next we perform an ANOVA using `aov`. The new feature is that we designate `subject` as a random factor with the `Error` term in our formula:

```
> aov.long.1 <- aov(y~1+age+Error(subject),data=dat.long.form)
> summary(aov.long.1) # p values assume sphericity
```

```

Error: subject
      Df Sum Sq Mean Sq F value Pr(>F)
Residuals 11  6624   602.2

Error: Within
      Df Sum Sq Mean Sq F value Pr(>F)
age      3    552  184.00  3.027 0.0432 *
Residuals 33  2006   60.79

---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

The resulting p value for age assumes sphericity. Note that no F test is performed on subjects.

11.6 association strength and effect size

Your textbook presents an equation for ω^2 , which represents the proportion of the variance of the treatment effects relative to the sum of all of the effects in the model:

$$\omega_A^2 = \frac{\sigma_\alpha^2}{\sigma_e^2 + \sigma_\pi^2 + \sigma_\alpha^2}$$

Another, perhaps more common index of the strength of association is partial-omega squared (Keppel and Wickens, 2004; Kirk, 1995), which expresses the variance of the treatment effects relative to the sum of only the error and treatment effects:

$$\omega_{Y|A.S}^2 = \frac{\sigma_\alpha^2}{\sigma_e^2 + \sigma_\alpha^2} \quad (7)$$

It can be calculated from the anova table with the using the formula:

$$\omega_{Y|A.S}^2 = \frac{(a-1)(F_A - 1)}{(a-1)(F_A - 1) + na} \quad (8)$$

where n is the number of subjects and a is the number of levels on variable A . For the data analyzed in the previous section, partial-omega squared for **age** is

$$\frac{3(3.0269 - 1)}{3(3.0269 - 1) + 48} = 0.112$$

Note that partial omega-squared is less than zero when $F_A < 1$; in such cases it is standard practice to set partial omega-squared to zero (Kirk, 1995). Partial omega squared can be used to derive an estimate of effect size, Cohen's f (Kirk, 1995):

$$\hat{f} = \sqrt{\frac{\omega_{Y|A.S}^2}{1 - \omega_{Y|A.S}^2}} \quad (9)$$

For our example, $\hat{f} = 0.35$.

11.7 linear comparisons among treatments

The strategy for conducting comparisons among treatments is similar to the one used previously for between-subjects designs. We start by creating a set of contrast weights that capture the comparison of interest: there should be one weight for each dependent measure, and all of the weights must sum to zero. Next, we create a composite score, ψ_i , for each subject that is simply the sum weighted dependent variables:

$$\psi_i = \sum_{j=1}^a c_j Y_{ij}$$

Finally, the values of the composite scores are evaluated using a `t.test`. If the null hypothesis begin evaluated by our comparison is non-directional, then we use `t.test` to test the null hypothesis that the composite scores were drawn from a zero-mean population. Consider, again, the data analyzed in section 11.5. Let us use a linear trend analysis to evaluate the hypothesis that `scores` followed a linear trend across `age`. The first step is to transform our data frame into a matrix of numbers:

```
> dat.mat <- with(mw115, cbind(age.30, age.36, age.42, age.48) )
> dat.mat
```

	age.30	age.36	age.42	age.48
[1,]	108	96	110	122
[2,]	103	117	127	133
[3,]	96	107	106	107
[4,]	84	85	92	99
[5,]	118	125	125	116
[6,]	110	107	96	91
[7,]	129	128	123	128
[8,]	90	84	101	113
[9,]	84	104	100	88
[10,]	96	100	103	105
[11,]	105	114	105	112
[12,]	113	117	132	130

The variable `dat.mat` represents the data as a four-column matrix: each row contains the data from one subject, and each column contains the data from a single age. Next, we create the contrast weights and then the composite scores using the matrix multiplication operator `%*%`. Note that the order of the terms is important here, so pay attention!

```
> lin.trend<-c(-1.5,-0.5,0.5,1.5);
> lin.scores<-dat.mat %*% lin.trend;
> lin.scores
```

```

      [,1]
[1,]  28
[2,]  50
[3,]  16
[4,]  26
[5,]  -3
[6,] -34
[7,]  -4
[8,]  43
[9,]   4
[10,] 15
[11,]  6
[12,] 33

```

Finally, we use `t.test` to evaluate the null hypothesis that the linear trend scores are drawn from a distribution with a mean of zero:

```

> t.test(lin.scores)

      One Sample t-test

data:  lin.scores
t = 2.2414, df = 11, p-value = 0.04659
alternative hypothesis: true mean is not equal to 0
95 percent confidence interval:
 0.2701827 29.7298173
sample estimates:
mean of x
      15

```

The t test is significant ($t = 2.24$, $df = 11$, $p = 0.046$) so we reject the null hypothesis that our composite scores, which represent the linear trend across age, are zero. Note that in this case the sphericity assumption *must* be valid because our comparison is a single degree of freedom test. Hence, there is a significant linear association between **score** and **age**.

It is also possible to do directional tests of our hypothesis. For example, here is how we could test the hypothesis that there is an *increasing* linear trend with age.

```

> t.test(lin.scores,alternative="greater")

      One Sample t-test

data:  lin.scores
t = 2.2414, df = 11, p-value = 0.0233
alternative hypothesis: true mean is greater than 0
95 percent confidence interval:
 2.981266      Inf

```

```
sample estimates:
mean of x
      15
```

The null hypothesis is that the composite scores are drawn from a population with a mean $\mu_{composite} \leq 0$; the alternative hypothesis is $\mu_{composite} > 0$. The test is significant, so we reject the null hypothesis in favor of the alternative. Note that the results of our directional test depends critically on the sign of the contrast scores:

```
> lin.trend<-c(1.5,0.5,-0.5,-1.5);
> lin.scores<-dat.mat%*%lin.trend;
> lin.scores

      [,1]
 [1,] -28
 [2,] -50
 [3,] -16
 [4,] -26
 [5,]  3
 [6,] 34
 [7,]  4
 [8,] -43
 [9,] -4
[10,] -15
[11,] -6
[12,] -33

> t.test(lin.scores,alternative="greater")

      One Sample t-test

data:  lin.scores
t = -2.2414, df = 11, p-value = 0.9767
alternative hypothesis: true mean is greater than 0
95 percent confidence interval:
 -27.01873      Inf
sample estimates:
mean of x
      -15
```

The bottom line is that you have to have a very clear understanding of how your contrast weights are related to your `t.test`.

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